

Fast model-free deconvolution of fluorescence decay for analysis of biological systems

Javier A. Jo

Qiyin Fang

Thanassis Papaioannou

Cedars-Sinai Medical Center

Department of Surgery

Biophotonics Research & Technology Development

Los Angeles, California 90048

Laura Marcu

Cedars-Sinai Medical Center

Department of Surgery

Biophotonics Research & Technology Development

Los Angeles, California 90048

and

University of Southern California

Departments of Electrical and Biomedical Engineering

Los Angeles, California

Abstract. For complex biological systems, conventional analysis of fluorescence intensity decay in terms of discrete exponential components cannot readily provide a true representation of the underlying fluorescence dynamics. We investigate an alternative nonparametric method for the analysis of time-resolved fluorescence data from biochemical and biological systems based on the expansion of fluorescence decay in a discrete Laguerre basis. We report that a unique Laguerre expansion can be found for fluorescence intensity decays of arbitrary form with convergence to a correct solution significantly faster than conventional multiexponential approximation methods. The Laguerre expansion coefficients are shown to be highly correlated with intrinsic fluorescence lifetimes and allow direct characterization of the fluorescence dynamics. A novel method for prediction of concentrations in mixtures of biochemical components using these coefficients is developed and successfully tested (prediction error $< 2\%$) using data from different mixtures of fluorescence lifetime standards. These findings suggest that the use of Laguerre expansion coefficients is a fast approach for the characterization and discrimination of complex biological systems such as tissues and cells, and that the method has potential for applications of fluorescence lifetime techniques to tissue diagnostics and imaging microscopy of living cells. © 2004 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.1752919]

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1 Introduction

Fluorescence spectroscopy is a nondestructive optical method extensively used to probe complex biological systems, including cells and tissues for biochemical, functional, and morphological changes associated with pathological conditions. Such an approach has potential for noninvasive diagnosis *in vivo*.^{1–3} Fluorescence measurements can be categorized as either static (steady state or time integrated) or dynamic (time-resolved). While steady state techniques provide an integrate spectrum over time that gives information about fluorescence emission intensity and spectral distribution, time-resolved techniques measure the dynamically evolving fluorescence emission, providing additional insight into the molecular species of the sample (e.g., the number of fluorescence species and their contribution to the overall emission) and/or changes in the local environment.^{2,3}

Two methods for time-resolved fluorescence measurements are widely used: The time-domain and the frequency-domain techniques. For the time-domain method, the sample is excited with a short pulse of light (typically nanosecond or shorter), and the emission intensity is measured following excitation with a fast photodetector. In the frequency domain, an

intensity-modulated light induces the sample fluorescence. Due to the fluorescence relaxation lifetime of the sample, the emitted wave is delayed in time relative to the excitation, inducing a phase-shift, which is used to calculate the decay time.³ The analytical approaches described here are applicable for analysis of time-domain data. The frequency-domain analysis is not discussed in this paper.

Analysis of time-resolved data from fluorescent systems includes determination of the intrinsic fluorescence intensity decay (fluorescence impulse response function, IRF), identification of a set of fitting parameters that best describe the characteristics of the fluorescence decay, and characterization and discrimination/classification of the fluorescent system based on those parameters.²

In the context of time-domain measurements, the fluorescence IRF contains all the temporal information of a single fluorescence decay measurement. The IRF is the system response to an ideal δ function excitation. In practice, the excitation light pulses are, typically, at least several picoseconds wide. Thus, they should be taken as a train of δ functions with different amplitudes; each one initiating an IRF from the sample, with an intensity proportional to the height of the δ function. The measured intensity decay function is the sum of all IRFs starting with different amplitudes and at different

Address all correspondence to Laura Marcu, Ph.D., Biophotonics Research Lab, Department of Surgery, 8700 Beverly Blvd., Davis Bldg. G149A, Cedars-Sinai Medical Center, Los Angeles, CA 90048. Tel: 310-423-8077; Fax: 310-423-8414; Email: lmarcu@bmsrs.usc.edu

times. Mathematically, the measured fluorescence intensity decay data is given by the convolution of the IRF with the excitation light pulse. Thus, to estimate the fluorescence IRF of a compound, the excitation light pulse must be deconvolved from the measured fluorescence intensity pulse.³

When the excitation light pulse is sufficiently short, resembling a δ function excitation, the measured fluorescence decay would closely resemble the intrinsic IRF.⁴ Although very short excitation light pulses can currently be generated using femtosecond lasers, due to their lack of general availability picosecond light sources are still the most commonly used for time-resolved measurements.³ Therefore, in many cases the intrinsic fluorescence IRF of the investigated compounds will have lifetimes on the order of the excitation light pulse width, and subsequently, an accurate deconvolution technique becomes crucial.

Deconvolution methods are usually divided into two groups⁵: those requiring an assumption of the functional form of the IRF, such as the nonlinear least-square iterative reconvolution method^{6,7} and those that directly give the IRF without any assumption, such as the Fourier⁸ and Laplace transform methods,⁹ the exponential series method,⁵ and the stretched exponential method,¹⁰ among others. In addition, an alternative approach known as global analysis, in which simultaneous analysis of multiple fluorescence decay experiments are performed, has proven useful for both time- and frequency-domain data.¹¹ Among these methods, however, the most commonly used deconvolution technique is the nonlinear least-squares iterative reconvolution (LSIR) method.⁵⁻⁷ This technique applies a least-squares minimization algorithm to compute the coefficients of a multiexponential expansion of the fluorescence decay. In complex biological systems, fluorescence emission typically originates from several endogenous fluorophores and is affected by light absorption and scattering. From such a complex medium, however, it is not entirely adequate to analyze the time-resolved fluorescence decay transient in terms of a multiexponential decay, since the parameters of a multiexponential fit cannot readily be interpreted in terms of fluorophore content.^{3,10} Moreover, different multiexponential expressions can reproduce experimental fluorescence decay data equally well, suggesting that for complex fluorescent systems there is an advantage in avoiding any *a priori* assumption about the functional form of the IRF decay physics.

Expansion on the discrete time Laguerre basis as means of deconvolving the intrinsic properties of a dynamic system from experimental input-output data was initially proposed by Marmarelis¹² and was applied to linear and nonlinear modeling of different physiological systems including renal autoregulation and autonomic control of heart rate.^{13,14} A Laguerre based deconvolution technique was recently reported as a variant of the LSIR technique, in which the fluorescence IRF is expressed as an expansion on the discrete time Laguerre basis instead of a weighted sum of exponential functions.¹⁵ The Laguerre deconvolution technique has been previously applied to optical spectroscopy of tissues with promising results to the analysis of time-resolved fluorescence emission data from arteriosclerotic lesions,^{1,2} and temporal spread functions of transmitted ultrafast laser pulses through different types of human breast tissue.¹⁶ However, a formal evaluation of this technique, as it applies to fluorescence measurements,

has not been reported. In this paper, the performance of the Laguerre deconvolution technique is analyzed in terms of accuracy and speed for retrieving the fluorescence IRF and compared to that of the multiexponential LSIR deconvolution technique.

Although the main objective of a deconvolution technique is to retrieve accurately the intrinsic fluorescence intensity decay characteristics of a compound, it is also very important that such a technique provides a unique representation of the intrinsic temporal fluorescence dynamics that allow further characterization of the investigated biological system. The study of Zacharakis et al.¹⁶ on the analysis of the temporal spread of transmitted ultrafast laser pulses through tissue represents the only attempt to directly use the Laguerre expansion coefficients to characterize biological systems, yielding promising but very preliminary results.¹⁶ Although the Laguerre deconvolution technique was used for the analysis of time-resolved fluorescence data from fluorescent constituents in tissues as well as tissue samples,^{1,2,15} the potential of Laguerre expansion coefficients for direct characterization and discrimination of fluorescence from biological systems has not been reported. Because these coefficients inherently contain information about the amplitude (intensity) and the relaxation time of a dynamic system, this paper investigates properties of Laguerre expansion coefficients that allow direct characterization of fluorescent system.

In summary, the goals of this paper were (1) to assess the performance of the Laguerre deconvolution technique in terms of its accuracy and speed for retrieving the intrinsic fluorescence decay from simulated and lifetime fluorescence standards data; (2) to investigate the analytical properties of the Laguerre expansion coefficients and to demonstrate their potential to directly characterize biochemical systems in terms of their fluorescence intensity decays; (3) to determine the potential use of Laguerre coefficients for quantitative interpretation of fluorescence data; and (4) to introduce a new method for prediction of concentration of mixtures of biochemical components that take advantage of the Laguerre coefficients information content.

2 Theoretical Approach

2.1 Laguerre Deconvolution Technique

The Laguerre deconvolution technique, in the context of time-domain time-resolved fluorescence emission data, is reviewed here. This nonparametric method expands the fluorescence IRF on the discrete time Laguerre basis.¹⁵ The Laguerre functions (LFs) have been suggested as an appropriate orthonormal basis owing to their built-in exponential term that makes them suitable for physical systems with asymptotically exponential relaxation dynamics.¹² Because the Laguerre basis is a complete orthonormal set of functions, a unique characteristic of this approach is that it can reconstruct a fluorescence response of arbitrary form. Thus, the Laguerre basis provides a unique and complete expansion of the decay function.

The measured fluorescence intensity decay data $y(t)$ is given by the convolution of the IRF $h(t)$ with the excitation light pulse $x(t)$:

$$y(t) = \int_0^t h(\tau)x(t-\tau) d\tau. \quad (1)$$

The time-domain time-resolved fluorescence measurements, however, are often obtained in discrete time, as in the case of pulse sampling and gated detection technique³ (direct recording of fluorescence pulse with a fast digitizer). In the discrete-time case, the relationship between the observed fluorescence intensity decay pulse and the excitation laser pulse is expressed by the convolution equation:

$$y(n) = T \sum_{m=0}^{K-1} h(m)x(n-m) \quad n=0, \dots, K-1. \quad (2)$$

The parameter K in Eq. (2) determines the extent of the system memory, T is the sampling interval, and $h(m)$ is the intrinsic fluorescence IRF. The Laguerre deconvolution technique uses the orthonormal set of discrete time LF $b_j^\alpha(n)$ to discretize and expand the fluorescence IRF:

$$h(n) = \sum_{j=0}^{L-1} c_j b_j^\alpha(n). \quad (3)$$

In Eq. (3), c_j are the unknown Laguerre expansion coefficients (LECs), which are to be estimated from the input-output data; $b_j^\alpha(n)$ denotes the j 'th order orthonormal discrete time LF; and L is the number of LFs used to model the IRF. The LF basis is defined as

$$b_j^\alpha(n) = \alpha^{(n-j)/2} (1-\alpha)^{1/2} \sum_{k=0}^j (-1)^k \binom{n}{k} \binom{j}{k} \alpha^{j-k} (1-\alpha)^k \quad n \geq 0. \quad (4)$$

The order j of each LF is equal to its number of zero-crossing (roots). The Laguerre parameter ($0 < \alpha < 1$) determines the rate of exponential decline of the LF. The higher the order j and/or the larger the Laguerre parameter α , the longer the spread over time of a LF and the larger the time separation between zero-crossing. Note that the Laguerre parameter α defines the time scale for which the Laguerre expansion of the system impulse response is most efficient in terms of convergence. Thus, fluorescence IRF with longer lifetime (longer memory) may require a larger α for efficient representation. Commonly, the parameter α is selected based on the kernel memory length K and the number of Laguerre functions L used for the expansion, so that all the functions decline sufficiently close to zero by the end of the impulse response.¹² This approach was applied in this work.

By inserting Eq. (3) into Eq. (2), the convolution Eq. (2) becomes

$$y(n) = \sum_{j=0}^{L-1} c_j v_j(n),$$

$$v_j(n) = T \sum_{m=0}^{K-1} b_j^\alpha(m)x(n-m), \quad (5)$$

where $v_j(n)$ are the discrete time convolutions of the excitation input with the LF and are denoted as the "key variables." The computation of the $v_j(n)$ can be accelerated significantly by use of the recursive relation:

$$v_j(n) = \sqrt{\alpha} v_j(n-1) + \sqrt{\alpha} v_{j-1}(n) - v_{j-1}(n-1), \quad (6)$$

which is due to the particular form of the discrete-time LF.¹² Computation of this recursive relation must be initialized by the following recursive equation that yields $v_0(n)$ for a given stimulus $x(n)$:

$$v_0(n) = \sqrt{\alpha} v_0(n-1) + T(1-\alpha)^{1/2} x(n). \quad (7)$$

This computation can be performed fast, for $n=0, 1, \dots, N$ and $j=0, 1, \dots, L-1$, where N is the number of samples in the data sets and L is the total number of LFs used in the IRF expansion. Finally, the unknown expansion coefficients can be estimated by generalized linear least-squares fitting of Eq. (5) using the discrete signals $y(n)$ and $v_j(n)$.

The optimal number of LFs and the value of the parameter α to be used in the model were determined by minimizing the weighted sum of the residuals:

$$S = \sum_{n=0}^{N-1} w_n [y(n) - \hat{y}(n)]^2, \quad (8)$$

where $y(n)$ is the real fluorescence decay, $\hat{y}(n)$ is the estimated decay, and w_n is the weighting factor. The weight w_n is proportional to the inverse of the experimental variance σ_n^2 for the measurements¹⁷ at time n . For time-correlated single-photon counting, which is the most common technique for time-domain measurements, it is straightforward to compute the experimental variance, since this is assumed to follow Poisson statistics, where the variance is known to be proportional to the number of photon counts.³ For other methods of time-domain measurements such as the direct recording of fluorescence pulse with a fast digitizer³ used in this paper, however, the experimental variance must be estimated from a representative set of experimental data. To estimate the experimental variance as a function of the amplitude of the fluorescence decay, repeated measurements of the sample fluorescence decay are taken, and the variance and the average intensity at each time point n of the decay signals are computed. The slope of the straight-line fit through a log-log plot of the variance as a function of the average intensity would indicate the relation between the experimental variance and the fluorescence intensity decay amplitude.¹⁷

In this study, the experimental variance was estimated from 55 repeated fluorescence decay measurements of 6 different fluorescence standard dyes at their peak wavelengths. The variance was represented in log-log scale as a function of the average of the 55 measurements. The average slope of the straight-line fits through the log-log plots for the six data sets was 0.99 (range: 0.91 to 1.09), suggesting that the experimental variance increased almost proportionally with the fluorescence signal. Thus, weight w_n was estimated by $1/y(n)$. For the case of the multiexponential LSIR technique, also applied to the data presented in this study, the weighted sum of the residuals of Eq. (8) was minimized to determine the coeffi-

ponents of a multiexponential expansion of the fluorescence decay. For purpose of clarification, the multiexponential model is defined by the following equation:

$$h(n) = \sum_{j=1}^P a_j \exp(-n/\tau_j), \quad (9)$$

where P is the number of exponentials chosen to represent the IRF $h(n)$, a_j are the preexponential factors, and τ_j are the time constants of the exponential terms.

2.2 Prediction of Concentration in Mixtures with the Laguerre Expansion Coefficients

Since the Laguerre expansion coefficients contain inherent information about the fluorescence amplitude (intensity) and the temporal decay characteristics, these coefficients can be directly used for quantitative analysis of the biochemical systems. To address this, a method for the prediction of concentrations in a mixture of biochemical components based on the analysis of the Laguerre expansion coefficients of the fluorescence IRF is introduced in this section and described as follows.

Let us assume that the sample fluorescence IRF $S(n)$ can be expanded on N Laguerre functions $b_j^\alpha(n)$:

$$S(n) = \sum_{j=0}^{N-1} c_j b_j^\alpha(n). \quad (10)$$

In Eq. (10), c_j are the expansion coefficients of the sample decay model. It is also assumed that the sample is composed of M biochemical components, each of them producing fluorescence IRF $C_k(n)$ that can also be expanded on the same N Laguerre functions:

$$C_k(n) = \sum_{j=0}^{N-1} a_{k,j} b_j^\alpha(n), \quad k=1, \dots, M. \quad (11)$$

In Eq. (11), $a_{k,j}$ are the expansion coefficients of the k 'th biochemical component IRF. Finally, we assume that the sample fluorescence IRF $S(n)$ can also be modeled as the linear combination of their M individual biochemical component fluorescence IRF $C_k(n)$:

$$S(n) = \sum_{k=1}^M A_k C_k(n), \quad (12)$$

where A_k are the relative contributions of the individual biochemical component fluorescence IRFs to the sample fluorescence IRF. Inserting Eq. (11) into Eq. (12), it can be followed that

$$\begin{aligned} S(n) &= \sum_{k=1}^M A_k C_k(n) = \sum_{k=1}^M A_k \left[\sum_{j=0}^{N-1} a_{k,j} b_j^\alpha(n) \right] \\ &= \sum_{j=0}^{N-1} \left(\sum_{k=1}^M A_k a_{k,j} \right) b_j^\alpha(n). \end{aligned} \quad (13)$$

Finally, from Eqs. (10) and (13), we can relate the expansion coefficients c_j of the sample IRF to the expansion coefficients $a_{k,j}$ of the biochemical component IRF as follows:

$$c_j = \sum_{k=1}^M A_k a_{k,j}, \quad j=1, \dots, N. \quad (14)$$

In practice, the expansion coefficients c_j and $a_{k,j}$ can be estimated from the sample fluorescence intensity decay and the individual biochemical fluorescence intensity decay measurements. Therefore, it is possible to retrieve the relative contributions A_k of the individual biochemical components fluorescence IRF to the sample fluorescence IRF, by solving the system of linear equations defined in Eq. (14). Notice that to solve this system, the number of equations should be greater than or equal to the number of unknowns ($N \geq M$); therefore, the number of LFs (N) used to expand the decay modeling should be greater than or equal to the number of the individual biochemical components (M).

The performances of the Laguerre deconvolution technique and the proposed method for the prediction of concentrations in a mixture of biochemical components based on the Laguerre expansion coefficients were assessed with simulated and experimental data. The radiative lifetime value τ_f of a given IRF was calculated by interpolating the time point at which the IRF becomes $1/e$ of its maximum value. A brief description of the simulated data generation, and of the fluorescence measurements on lifetime fluorescence standards is presented in the next section.

3 Generation of Time-Resolved Fluorescence Data

3.1 Simulated Data

The synthetic data was generated by means of a four-exponential model, with fixed decay constants and random values for the fractional contribution of each exponential component. A total of 600 data sets were generated, yielding average lifetime values between 0.3 and 12 ns. White noise of zero mean and three different variance levels was added to the data, yielding three different groups of 600 data sets, with approximately 80, 60, and 50 dB SNRs (low-noise-, medium-noise-, and high-noise-level groups, respectively). A laser pulse (700-ps pulse width) measured from a sample of 9-cyanoanthracene (see next section) was used as the excitation signal for our simulation. Therefore, all the data sets were convolved with the laser signal, yielding the "measured" decay data for our simulation. The Laguerre deconvolution technique was applied to this data, using different model orders ranging from three to six Laguerre functions. Similarly, the multiexponential deconvolution technique was also applied to the data, using different model orders ranging from one to four exponential components.

3.2 Experimental Data: Lifetime Fluorescence Standards

Data were collected from standard dyes for fluorescence lifetime measurements. The dyes were selected to cover a broad range of radiative lifetimes (0.54 to 12 ns) that are most relevant for biological applications, such as fluorescent emission from tissue. The fluorophores chosen included rose bengal (33,000, Sigma-Aldrich), rhodamin B (25,242, Sigma-Aldrich), and 9-cyanoanthracene (15,276, Sigma-Aldrich). The fluorescence dyes used in the measurements were diluted

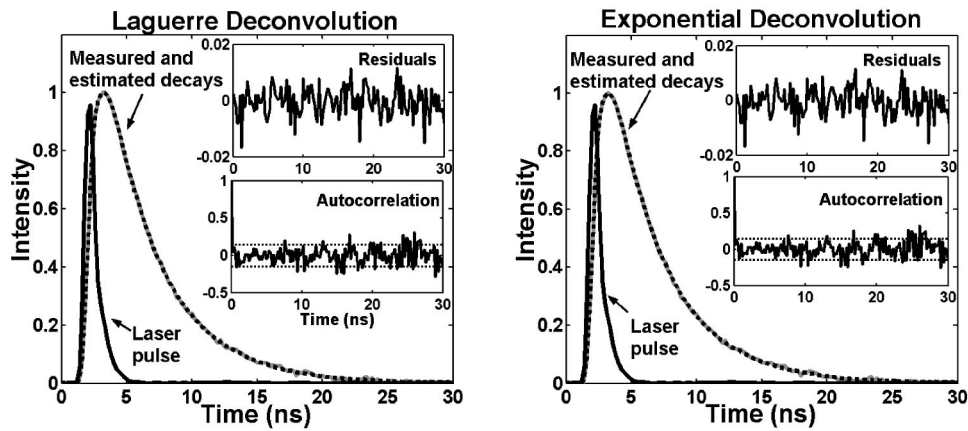


Fig. 1 Laguerre and multiexponential deconvolution of the excitation laser pulse (solid black) from a simulated fluorescence decay (solid gray, $\tau = 4.1$ ns). The estimated fluorescence decays (dotted black) overlap the simulated fluorescence decay. The residuals and their autocorrelation function indicate excellent fit between the synthetic and the estimated fluorescence decays.

into 10^{-6} M solutions. The fluorescence standard samples were excited with a subnanosecond pulsed nitrogen laser with an emission wavelength of 337.1 nm (700 ps FWHM). The fluorescence response was measured using a time-resolved time-domain fluorescence apparatus, enabling direct recording of the fluorescent pulse (fast digitizer and gated detection). The fluorescence pulse was collected by a fiber optic bundle (bifurcated probe) and directed to a monochromator connected to a multichannel plate photomultiplier tube with a rise time of 180 ps. The entire fluorescence pulse from a single excitation pulse was recorded with a 1-GHz bandwidth digital oscilloscope.¹⁸ For each sample solution, the time-resolved fluorescence spectra were measured for a 250-nm spectral range from 400 to 650 nm at 5-nm increments. After each measurement sequence, the laser pulse temporal profile was measured at a wavelength slightly below the excitation laser line. Background spectra were taken for the solvents (ethanol or methanol) using the same cuvette.

The method for prediction of concentrations in a mixture of biochemical components based on the analysis of the Laguerre expansion coefficients was tested on mixtures of rose bengal (RB) and rhodamin B (RdmB) of distinct relative concentrations. Three types of mixture solutions were prepared with $[RdmB]/[RB]$ concentration values equal to 0.25/0.75, 0.5/0.5, and 0.75/0.25 μM , respectively. The 10^{-6} M RB and RdmB solutions were also measured representing $[RdmB]/[RB]$ concentrations of 0/1.0 and 1.0/0 μM , respectively. Time-resolved fluorescence measurements at wavelengths between 550 and 600 nm (corresponding to the range of wavelengths around the spectral peak at 575 nm) were recorded from the five solutions and used for the analysis.

4 Technique Evaluation and Validation

4.1 Simulated Data

A simulated time-resolved fluorescence intensity decay ($\tau = 4.1$ ns at a medium noise level), its corresponding estimation by the Laguerre deconvolution method ($L=5$), and the laser pulse (solid black) are shown in Fig. 1. The residuals are $<2\%$ of the peak fluorescence amplitude and they appear randomly distributed around zero. The autocorrelation func-

tion of the residuals does not contain low-frequency oscillations characteristic of nonrandom residuals, and is mostly contained within the 95% confidence interval centered around zero. These observations indicate an excellent fit between the synthetic and estimated fluorescence decays, showing that the fluorescence IRF was properly estimated with the deconvolution algorithm based on the Laguerre expansion technique. The results for the same sample fluorescence decay using the multiexponential approach ($P=3$), presented in Fig. 1, showed similar results. One important detail depicted in Fig. 1 is that the residuals and autocorrelation functions corresponding to both the Laguerre and multiexponential fits look very much alike. This is explained by the fact that both techniques were able to accurately fit the true synthetic time-resolved decay data, leaving out just the additive white noise component of the artificial data.

The performance of Laguerre and multiexponential deconvolution techniques along the lifetime range of 0.3 to 12 ns was assessed by means of the relative error between the real and the estimated lifetime (RLE) values, and the normalized mean square error (NMSE). Similar performances were obtained for both techniques: the RLE values were found below 1, 2, and 4%, whereas the NMSE values were smaller than 0.04, 0.3, and 0.9% for the low-, medium-, and high-noise-level groups, respectively.

To investigate the effect of the number of LF (chosen to expand the fluorescence IRF) on the estimation of the intrinsic fluorescence decay, the synthetic data was deconvolved using Laguerre expansions of different orders (three to six LFs). For decays with lifetimes ranging from 1 to 8 ns, the IRF expansion with five LF yielded the best estimation of the fluorescence emission decay ($RLE < 2\%$). The best estimate for fast decays ($\tau_f < 1$ ns) and for slow decays ($\tau_f > 8$ ns) were obtained using an expansion of six LFs and three to four LFs, respectively. For the multiexponential deconvolution method, a good estimation of the IRF ($RLE < 2\%$) was yielded by biexponential expansion for slow decays and triexponential expansion for fast decays.

As stated, it is important that a deconvolution technique provides a representation of the intrinsic temporal fluorescence dynamics to be used for further characterization of the

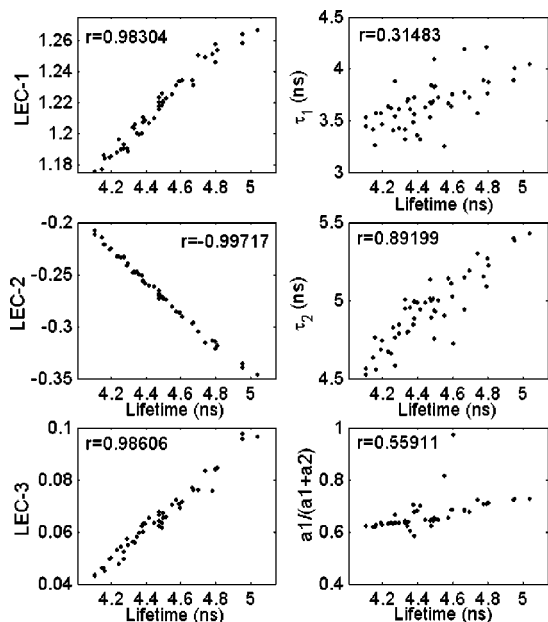


Fig. 2 Correlation between the actual lifetime values of the simulated data with the Laguerre expansion coefficients (LEC-1 to LEC-3, left panels) and the multiexponential LSIR parameters (right panels).

investigated compound. Both the multiexponential and Laguerre deconvolution techniques summarize the temporal fluorescence intensity decay information in terms of the parameters of the model they use to represent the fluorescence IRF: (1) the preexponential factors (a_i) and the decay constants (τ_i) for the case of the multiexponential approach and (2) the Laguerre α parameter and the c_j expansion coefficients for the case of Laguerre approach. Therefore, a natural attempt to characterize a compound in terms of its fluorescence lifetime information is to utilize either the multiexponential or the Laguerre model parameters.

To investigate whether these model parameters reflect by themselves the fluorescence temporal information of the investigated compound, the correlation coefficients between the actual lifetime values of the simulated data and the model parameters (first two exponential parameters and first three Laguerre expansion coefficients) were computed. For a bivariate case, the correlation coefficient ($-1 < r < 1$) is defined as

the covariance between the two variables normalized by the variances of each variable, and measures the strength of the linear relationship between the two variables. A correlation coefficient $r = -1$ indicates a perfect negative (inverse) linear dependence, $r = 0$ indicates no linear dependence, and $r = 1$ indicates a perfect positive linear dependence.⁴

The first three Laguerre expansion coefficients as a function of the radiative lifetime between 4 and 5 ns and the correlation coefficients are shown in Fig. 2. We can clearly observe the first and third expansion coefficients were positively correlated with the intrinsic decay lifetime, whereas the second expansion coefficient was negatively correlated with the intrinsic lifetimes. All three Laguerre expansion coefficients (LEC-1 to LEC-3) were highly correlated to the real lifetime values ($r > 0.95$). This result indicates that each of the Laguerre expansion coefficients capture and reflect the temporal relaxation of the IRF, and thus can be further used for the characterization of the investigated compound.

Plots of the biexponential fast (τ_1) and slow (τ_2) decay constants and of the relative contribution of the fast exponential [$a_1/(a_1 + a_2)$] as a function of the radiative lifetime between 4 and 5 ns are also shown in Fig. 2. For this particular lifetime range, only one of the time-decay constants, the slow decay constant (τ_2), was positively correlated ($r = 0.82 \pm 0.01$) with the lifetime values. The remaining time decay constant and the normalized preexponential factors were not correlated with the lifetime values ($r = 0.55 \pm 0.02$ and $r = 0.54 \pm 0.03$, respectively).

4.2 Experimental Data

To investigate the accuracy of the Laguerre deconvolution method for retrieving fluorescence decays from experimental data, a number of lifetime fluorescence standards with a broad range of relaxation lifetimes were measured in solutions. The lifetimes retrieved using both the Laguerre and the multiexponential deconvolution methods were compared with values from the literature.³ The results of this analysis are presented in Table 1 [mean \pm standard error (SE)]. Short-lived fluorophores, such as rose bengal, with lifetimes ranging in the hundreds of picoseconds could be reliably retrieved by the Laguerre deconvolution technique. This is shown in Fig. 3 for a fluorescence intensity decay measurement from RB in methanol at 580 nm. The fluorophore intensity decay data was best fitted to Laguerre models of $L = 3$ or $L = 4$ and to a single-

Table 1 Lifetime values (mean \pm SE) of fluorescence standard dyes estimated using the Laguerre and exponential deconvolution techniques.

Sample	Solvent	Lifetime (ns)			Wavelength (nm)
		Laguerre	Exponential	Literature ³	
RB	Ethanol	0.769 \pm 0.005	0.702 \pm 0.004	0.850	590
RB	Methanol	0.457 \pm 0.004	0.400 \pm 0.006	0.540	580
RdB	Ethanol	2.872 \pm 0.015	2.880 \pm 0.006	2.850	590
RdB	Deionized water	1.533 \pm 0.057	1.531 \pm 0.024	1.520	580
9/CA	Ethanol	11.746 \pm 0.026	11.630 \pm 0.020	11.850	445

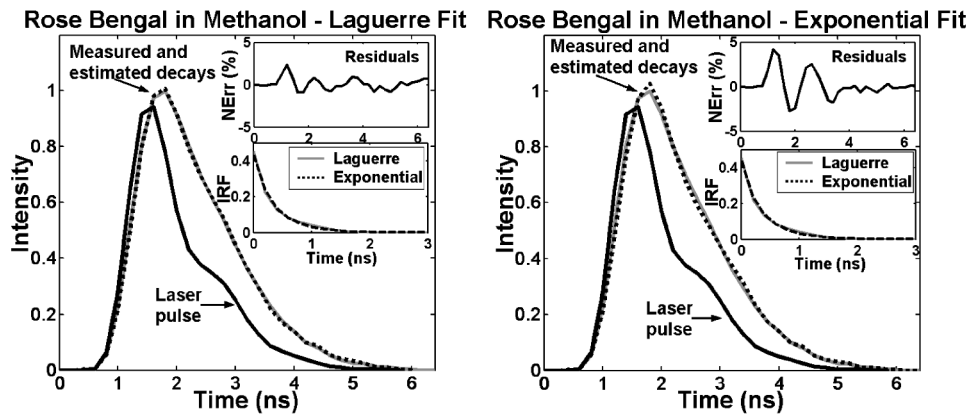


Fig. 3 Laguerre and multiexponential deconvolution of the excitation laser pulse (solid black) from a measured fluorescence decay from RB in methanol at 580 nm (solid gray). The estimated fluorescence decays (dotted black) overlap the measured fluorescence decay. The residuals indicate excellent fit between the measured and the estimated fluorescence decays. The estimated IRF using both methods present very similar decay characteristics.

exponential decay. Both methods yielded very good fits, although in this particular example, the Laguerre approach performed better than the multiexponential method, as indicated by the smaller Laguerre residuals (top insets). Both the Laguerre and the exponential methods yielded similar intrinsic fluorescence decays (IRF) with lifetime values of 0.456 ± 0.004 and 0.4 ± 0.006 ns, respectively, which were in good agreement with previously reported data.³

Similarly, measurements of RdmB in ethanol and methanol demonstrated the ability of the Laguerre method to accurately resolve nanosecond and subnanosecond fluorescence lifetimes (Fig. 4), which is of especial importance since a number of biologically relevant fluorophores such as elastin and collagen are known to emit at these time scales.^{2,15} The RdmB fluorophore intensity decay data was best fitted to Laguerre models of $L=4$ and to a single exponential decay. Both methods yielded very good fits as it can be seen in Fig. 4.

Finally, to assess the ability of the Laguerre technique to retrieve long fluorescence lifetimes, the fluorescence decay of 9-cyanoanthracene (9CA) in ethanol were also deconvolved

(Fig. 5). Both the Laguerre and the exponential methods yielded similar intrinsic fluorescence decays with lifetime values (Table 1) also in agreement with previous reports.³ The 9CA fluorophore intensity decay data was best fitted to Laguerre models of $L=2$ and to a single exponential decay, and very good fits were attained by the two methods as observed in Fig. 5. Similar to the results on the simulation data, it was observed that deconvolution of long-lived fluorophores (e.g., 9CA) require fewer LFs for expansions when compared with the short-lived compounds (e.g., RB and RdmB).

To compare the computational time of both the Laguerre and the multiexponential deconvolution techniques, 250 sets of 9CA time-resolved fluorescence data at several wavelengths (400 to 650 nm) were deconvolved using both techniques, and the deconvolution time was measured for each set of data. The average computational time was 33.1 ± 0.86 ms for the Laguerre deconvolution with five LFs, and 101.4 ± 1.6 ms for the LSIR deconvolution with a single exponential. Both methods were implemented in Matlab-6.5 (The

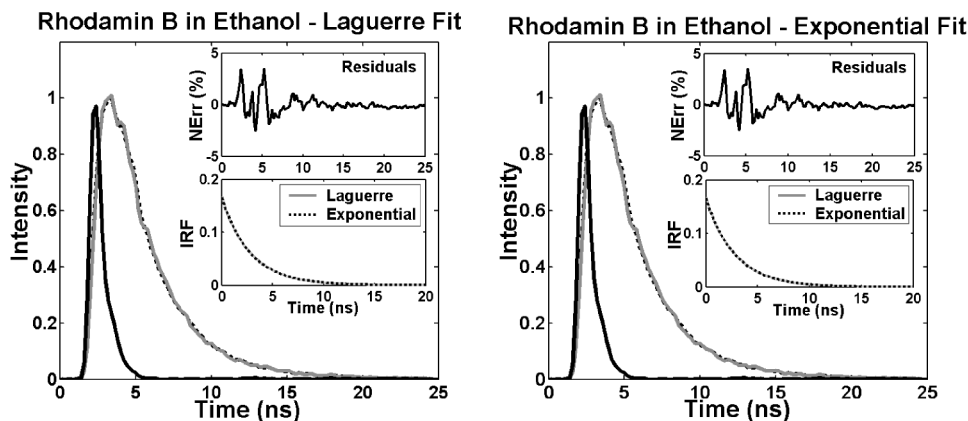


Fig. 4 Laguerre and multiexponential deconvolution of the excitation laser pulse (solid black) from a measured fluorescence decay from RdmB in ethanol at 590 nm (solid gray). The estimated fluorescence decays (dotted black) overlap the measured fluorescence decay. The residuals indicate excellent fit between the measured and the estimated fluorescence decays. The estimated IRF using both methods present very similar decay characteristics.

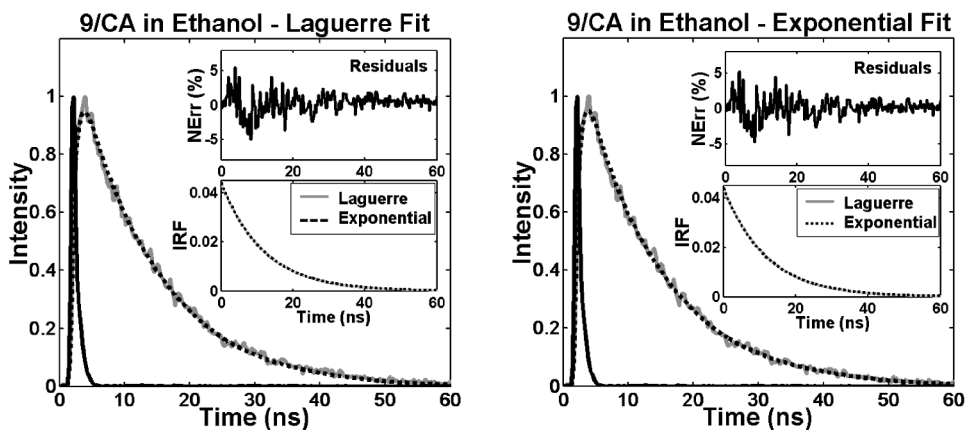


Fig. 5 Laguerre and multiexponential deconvolution of the excitation laser pulse (solid black) from a measured fluorescence decay from 9/CA in ethanol at 445 nm (solid gray). The estimated fluorescence decays (dotted black) overlap the measured fluorescence decay. The residuals indicate excellent fit between the measured and the estimated fluorescence decays. The estimated IRF using both methods present very similar decay characteristics.

MathWorks, Natick, MA) and executed on an IBM-compatible workstation (Intel Xeon processor, 2.0 GHz).

4.3 Concentration Prediction with Laguerre Coefficients

To test the proposed method for the prediction of concentrations in a mixture of biochemical components, fluorescence decays of five mixtures of RdmB and RB were expanded using five LFs with discrete-time Laguerre parameter $\alpha = 0.81$. The relative contributions (A_k) of the individual biochemical component decays to the mixture fluorescence decays were predicted by solving Eq. (14). To compare the proposed approach with more traditional methods for concentration prediction based on spectral analysis, the relative concentration of RdmB was also determined by applying¹⁹⁻²¹ principal component regression (PCR) and partial least squares (PLS) to the spectral data of the mixtures. The results of the three methods are shown in Table 2. All methods give a close estimation of the RdmB concentration in the three solutions. However, the Laguerre model of intrinsic fluorescence decays yielded a better estimation of the fluorophore concentrations (error < 2%), compared to the spectral methods (PCR: error < 7%; PLS: error < 10%).

Table 2 Predicted RdmB concentrations from the three solutions estimated by the three methods applied.

Method	Relative Concentration of RdmB		
	25%	50%	75%
PCR	22.8 ± 1.25	48.79 ± 1.77	68.66 ± 1.57
PLS	21.98 ± 1.63	47.31 ± 1.99	67.04 ± 1.69
Laguerre	25.23 ± 0.48	51.16 ± 0.83	73.86 ± 1.66

5 Discussion

Although a multiexponential deconvolution has the potential to accurately retrieve the fluorescence IRF of complex biomolecular systems, the parameters of a multiexponential fit cannot readily be interpreted in terms of number of fluorophore content.^{3,5} Changes of a fluorophore environment, proteins conformations or cross-links, also would result in different intensity decay for a single fluorophore,^{3,5} thus it is not practical to consider individual decay times. Moreover, very different multiexponential expressions can reproduce the same experimental fluorescence intensity decay data equally well as demonstrated with the simulation results presented here, in which synthetic decay data generated by a four-exponential model was accurately deconvolved by multiexponential models of different orders from two to four exponential components. This and previous reported evidence^{3,5,10} support the conclusion that for complex fluorescence systems, there is an advantage in avoiding any assumption about the functional form of the fluorescence decay law. Further, this suggests that the Laguerre deconvolution technique is a suitable approach for the analysis of time-domain fluorescence data of complex systems, since this technique has the ability to expand intrinsic fluorescence intensity decays of any form, without any *a priori* assumption of its functional form.

While the results from both simulation and lifetime fluorescence standard support the conclusion that the Laguerre method performs similarly to the multiexponential method in terms of accuracy for retrieving the fluorescence IRF, one advantage of the Laguerre method is that for any value of the Laguerre parameter α , the corresponding basis of LF is a complete orthonormal basis. Thus, it is certain that an expansion of the fluorescence IRF on the LF basis can always be found, and even more important, the set of expansion coefficients are unique for a defined Laguerre basis. Our results showed that three to six LFs are sufficient to expand fluorescence decays with lifetime values ranging from 0.3 to 12 ns, which are times relevant for the emission of tissue endogenous fluorophores. Generally, an accurate estimation of slow

decays requires fewer LFs for expansion (less than three) than for the fast decays (larger than five). In contrast, deconvolution with the multiexponential approach may yield more than one solution, even when the number of exponential or the values of the decay constants are prefixed. The span of possible solutions of a multiexponential expansion can be significantly reduced by fixing *a priori* the values of the decay constants, as was proposed in the exponential series method proposed by Ware et al.⁵ Note, however, that deconvolution with the exponential series method is successful only when the prefixed decay constants are commensurate with the investigated fluorescence decay.

Another advantage of the Laguerre deconvolution over the multiexponential LSIR results from the different methods required to estimate the expansion parameters. The Laguerre expansion technique uses a least-squares optimization procedure to determine the coefficients of the Laguerre expansion of the system dynamics.¹² For a defined Laguerre basis, i.e., given values of the parameter α and the number of the LFs, the problem of finding the expansion coefficients is reduced to solving an overdetermined system of linear equations [given in Eq. (5)], which represents a linear least-squares minimization problem. The same property holds even for the estimation of nonlinear dynamics that can also be formulated in terms of linear equations using the Laguerre expansion technique.¹²⁻¹⁴ In contrast, traditional multiexponential LSIR techniques require the estimation of intrinsic nonlinear parameters (the decay constants), which represents a nonlinear least squares problem. Although very robust and efficient nonlinear least square algorithms are currently available, such as the Gauss-Newton and Levenberg-Marquardt methods,²² solving a linear least-squares problem (Laguerre technique) is a much simpler and less computationally expensive problem than finding a nonlinear least square solution (through a multiexponential technique). This was clearly supported by the results described here on the speed of computational analysis, showing that the Laguerre deconvolution technique converges to a correct solution approximately three times faster than the monoexponential deconvolution. The convergence speed would become more significant when a biexponential or higher order multiexponential expansion is used. This specific advantage of the Laguerre method becomes even more important in the context of application of lifetime fluorescence spectroscopy to clinical research of tissue diagnosis, where the speed of data analysis is of crucial importance.

Analysis of correlations coefficients (r) demonstrated that each Laguerre expansion coefficient is highly correlated with the intrinsic lifetime value, suggesting that the use of these coefficients has potential, as a new approach, for the direct characterization of biochemical compounds in terms of their fluorescence emission temporal properties. For the case of the multiexponential deconvolution, although the estimated (computed) average lifetime is always correlated with the intrinsic radiative lifetime, the individual multiexponential parameters (decay constants and preexponential coefficients) may not necessarily be correlated to the intrinsic lifetimes. This was shown in the simulations results, where only one of the decay constants was correlated with the lifetime. This lack of correlation between individual multiexponential parameters and the lifetime occurs because the multiexponential model does not represent an orthogonal expansion of the fluorescence IRF;

therefore, the estimated fitting parameters are not independent from each other.^{3,23} This condition implies that the value of one specific parameter is determined not only by the data to be fitted, but also by the value of the other fitting parameters; thus, reducing the correlation between the actual lifetimes and the estimated values of the multiexponential parameters. In contrast, since the Laguerre basis provides an orthogonal expansion of the IRF, the value of each expansion coefficient depends exclusively on the data to be fitted, making them highly correlated to the actual lifetime values.

This paper also demonstrated that the Laguerre expansion coefficients have potential for quantitative interpretation of fluorescence decay. A new method for the prediction of concentrations in a mixture was introduced here and successfully tested on fluorescence standards components (RdmB and RB). Moreover, using only fluorescence decay information from a narrow spectral range, this method yielded improved prediction of the fluorophore concentrations when compared to that obtained using traditional methods of spectral analysis (PCR and PLS). One possible explanation for this result is that spectral methods use the information derived from spectral distribution of fluorescence intensity, while the proposed Laguerre method uses both the amplitude and temporal information of the fluorescence emission from a predefined narrow spectral range. Emission spectra of RdmB and RB are highly overlapped,³ thus spectral emission alone would provide limited information for the prediction of their relative concentration. However, RdmB and RB present different fluorescence decay characteristics, which were taken into account together with the amplitude information when the Laguerre coefficients were applied for concentration prediction. Although in this paper, the proposed Laguerre method analyzes only the amplitude and temporal information of the fluorescence emission from a predefined narrow spectral range, the Laguerre expansion coefficients corresponding to fluorescence IRF taken at multiple emission wavelengths also reflect the spectral information of the fluorescence emission. This spectral information could also be integrated into a more advanced method for characterization and discrimination of a biological system.

It is also noteworthy that an analogous method for the prediction of concentrations in a mixture of biochemical components based on the multiexponential model of intrinsic fluorescence decay could also be implemented. Since such a method would also use both the amplitude and temporal information contained by fluorescence data, it would most likely yield a performance similar to that of the Laguerre method presented in this paper. However, a method based on the multiexponential expansion would require that the decay constants be chosen *a priori*. Therefore, one disadvantage of this approach would represent the necessity for a good guess of the decay constants (these must be commensurate with the investigated fluorescence decays) to ensure an acceptable prediction of the investigated concentrations.⁵ A further requirement for such a technique would be that the number of exponential terms used should be equal to or greater than the number of biochemical concentrations to be retrieved. Therefore, the larger the number of component assumed to be present in the investigated biological system, the more exponential terms are required, and a large number of decay constants must be guessed, making its application difficult. The

Laguerre-model-based method for concentration estimation requires only predefining the Laguerre parameter α , which can be chosen from the observed decay data.¹²

All these findings taken together suggest that the application of the Laguerre expansion method (and specifically, of the expansion coefficients) represent a promising approach for the quantification of relative concentrations of different biochemical compounds in biological systems, and their subsequent characterization and discrimination. Application of the Laguerre deconvolution techniques can result not only in accurate analysis of fluorescent systems without *a priori* knowledge of underlying fluorescence dynamics but also facilitates reduction of data processing time. This method can be easily adapted for the analysis of time-resolved fluorescence data from tissues, and thus it has potential impact on applications of fluorescence lifetime to *in vivo* spectroscopic diagnostics of diseased tissues (e.g., arterial plaques and tumors) and functional imaging microscopy of living cells.

In conclusion, this work demonstrated that numerical deconvolution of fluorescence IRF using a nonparametric expansion in a discrete-time Laguerre basis enables accurate estimation of the intrinsic fluorescence intensity decays for a broad range of lifetime values. Although the main objective of a reliable deconvolution technique is to accurately retrieve the intrinsic fluorescence decay of a compound, it would also be very important for such technique to provide a unique representation of the intrinsic fluorescence dynamics that can be used for further characterization of the investigated biological system. Taking into account this consideration, our results demonstrated that the Laguerre deconvolution method includes a number of interesting properties and potential advantages over the classical multiexponential LSIR. These include faster convergence to a correct solution, high correlation between the expansion coefficients and the intrinsic lifetime values, and the potential for directly quantifying the relative concentrations of different biochemical compounds in biological systems. These characteristics suggest that the use of the Laguerre expansion coefficients represents a new reliable and powerful approach for the characterization and discrimination of biological systems, in terms of their fluorescence emission intensity amplitude and temporal or lifetime characteristics. Future work will investigate the application of this approach to analysis of time-resolved fluorescence data obtained from *in vivo* measurements in diseased tissue including atherosclerotic lesions and brain tumors.

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